

*Functional DNA architectures:
Photoinduced electron transfer and
switchable optical properties*



Hans-Achim Wagenknecht

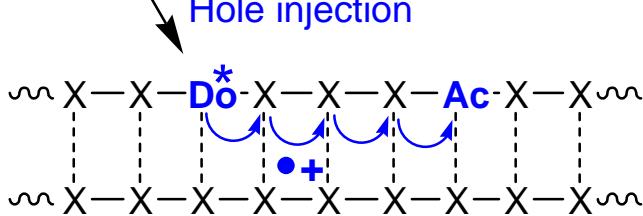
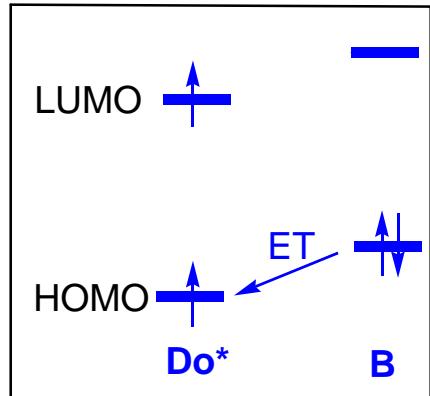


Universität Regensburg

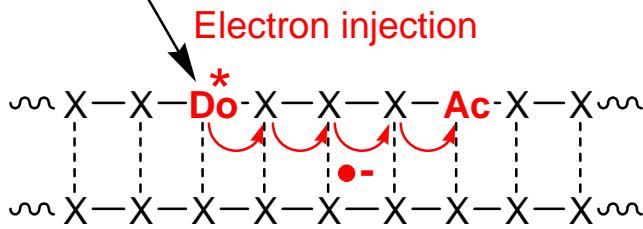
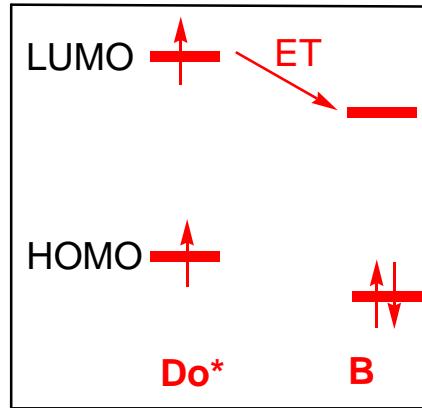
Part I+II: Photoinduced charge transfer in DNA



Hole transfer vs. electron transfer



- Superexchange
- Hopping

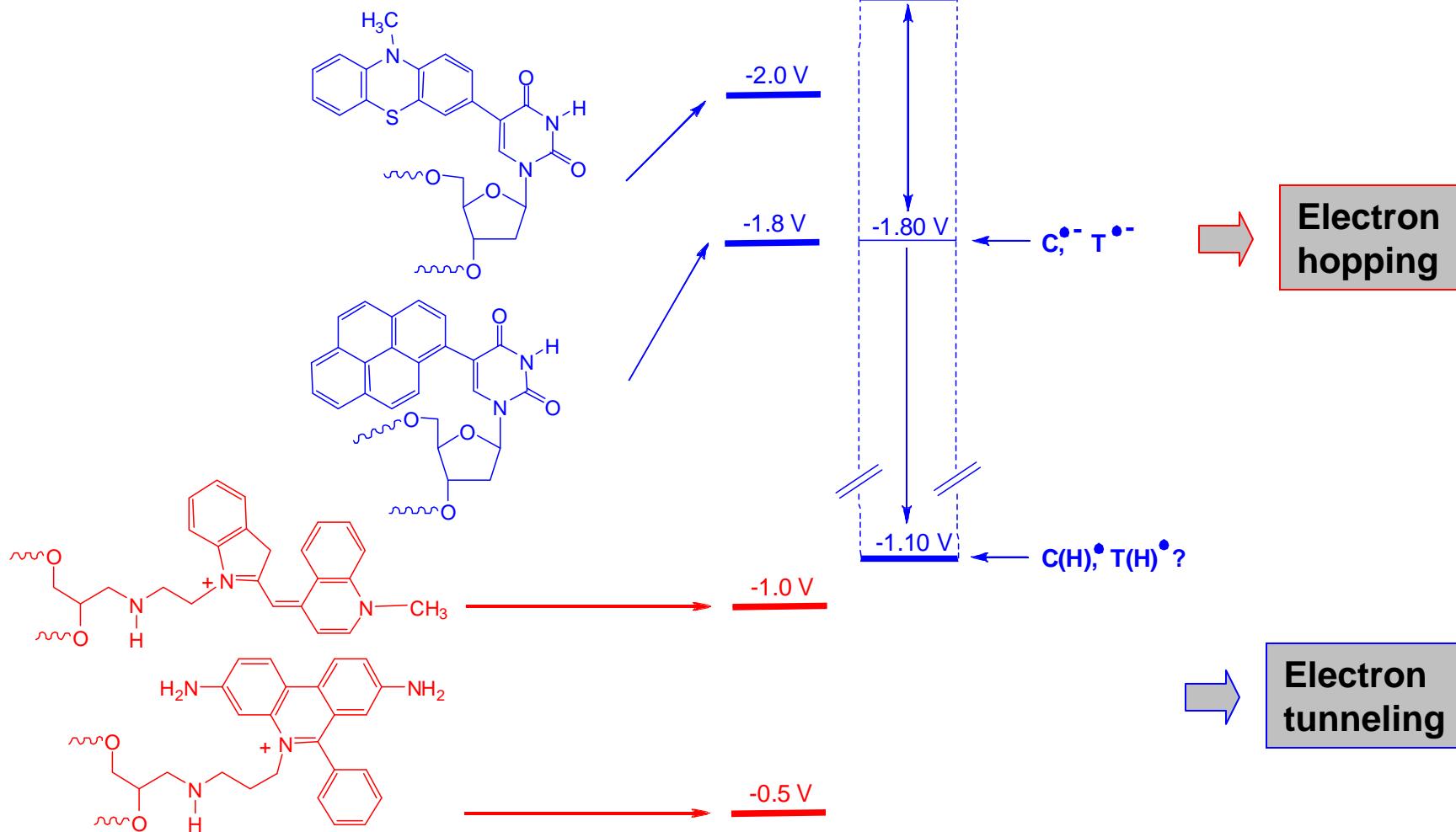


Proposed to be
more suitable for nano/biotechnology:

- faster
- more efficiently
- less (or no) damage

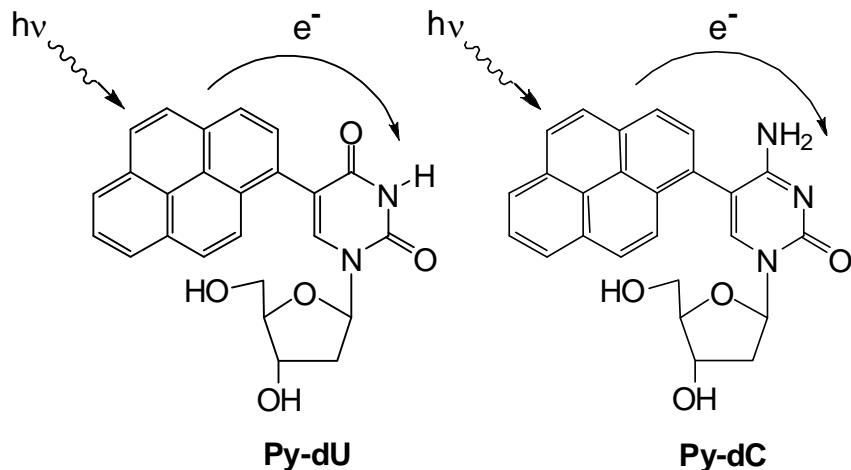
Part I+II: Photoinduced electron transfer in DNA

Excited state potentials of electron donors



Part I. Nucleoside models for electron transfer in DNA

Summary



k_{ET} : 4.7 ps
 $pK_a = 5.5$

k_{ET} : 40 ps
 $pK_a > 12$

Redox potentials:

C and T as electron carriers in DNA



Nucleoside model studies:

- T is reduced faster
- $d\text{C}^{\bullet-}$ exhibits strong basicity



Implication for DNA:

$d\text{T}^{\bullet-}$ is the major electron carrier

Synlett 2002, 687.

Angew. Chem. Int. Ed. 2002, 41, 2978.

Chem. Commun. 2003, 1878 .

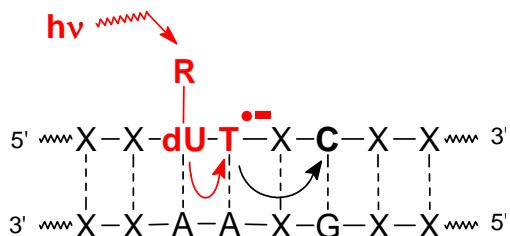
Synthesis 2003, 2335.

ChemPhysChem 2004, 5, 706

Chem. Phys. Lett. 2005, 409, 277.

Nicole Amann, Elke Mayer-Enhart
In collaboration with Torsten Fiebig, Boston

Electron injection studies



Chem. Eur. J. **2002**, *8*, 4877-4883.

Eur. J. Org. Chem. **2003**, 2498.

Angew. Chem. Int. Ed. **2004**, *43*, 1845.

DNA studies

Angew. Chem. Int. Ed. **2003**, *42*, 2454.

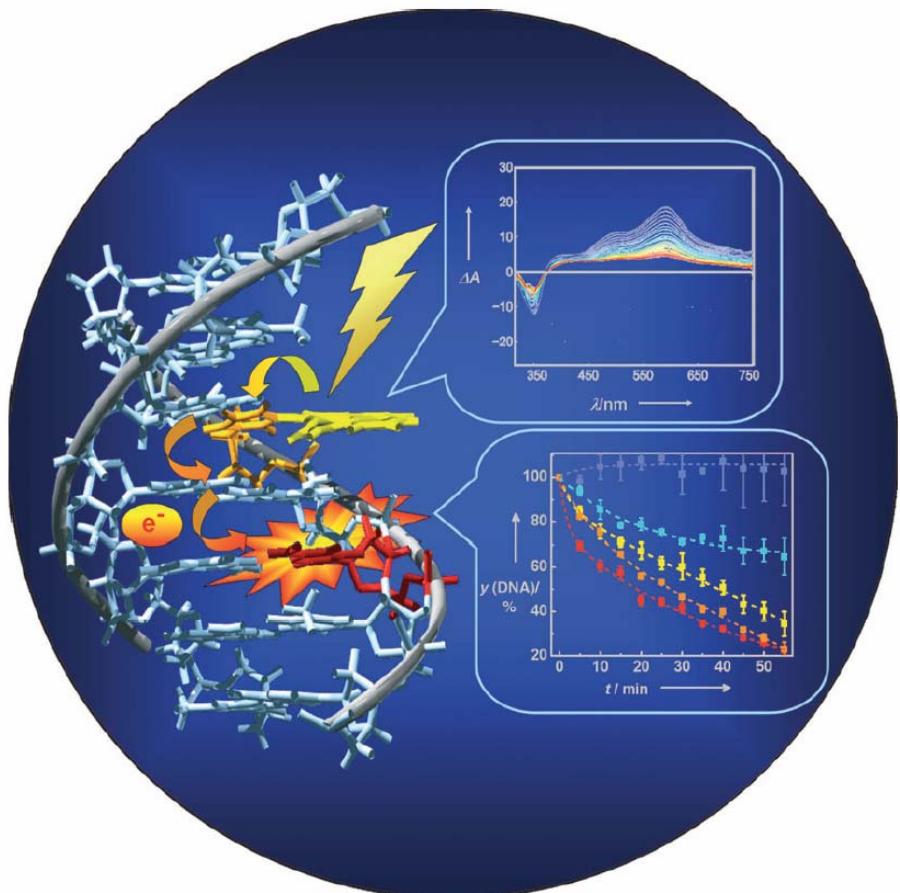
Chem. Eur. J. **2005**, *22*, 1871.

Angew. Chem. Int. Ed. **2005**, *44*, 1636.

Proc. Natl. Acad. Sci. USA **2006**, *103*, 10192

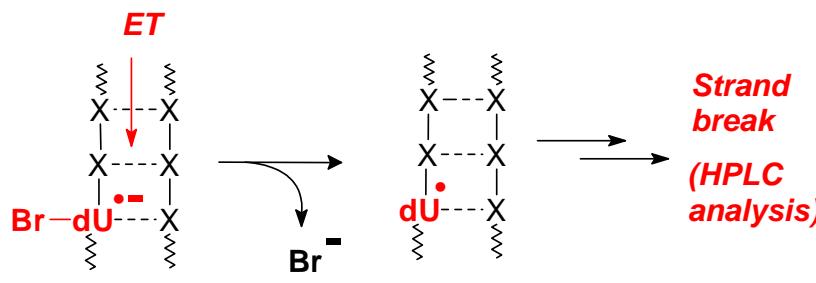
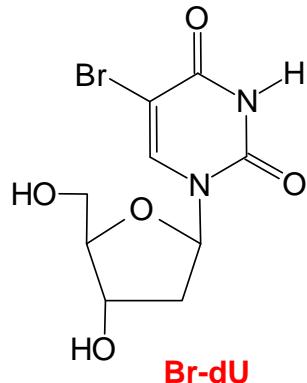
Communications

*Angewandte
Chemie*



The reductive electron transfer (ET) in DNA can be studied by ultrafast time-resolved measurements combined with chemically probed DNA-strand-cleavage experiments. Owing to the numerous conformations of DNA present the results show a variety of ET rates. For more information see the Communication by H.-A. Wagenknecht, T. Fiebig, et al. on the following pages.

Chemical electron acceptor



DNA studies

Angew. Chem. Int. Ed. **2003**, *42*, 2454.

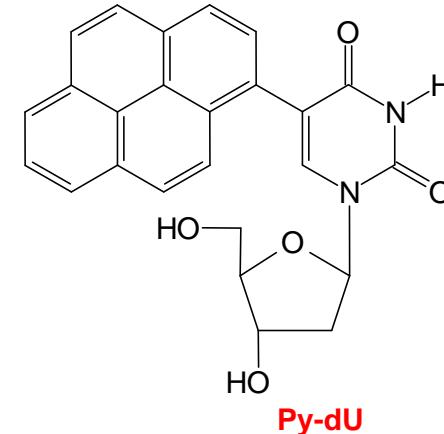
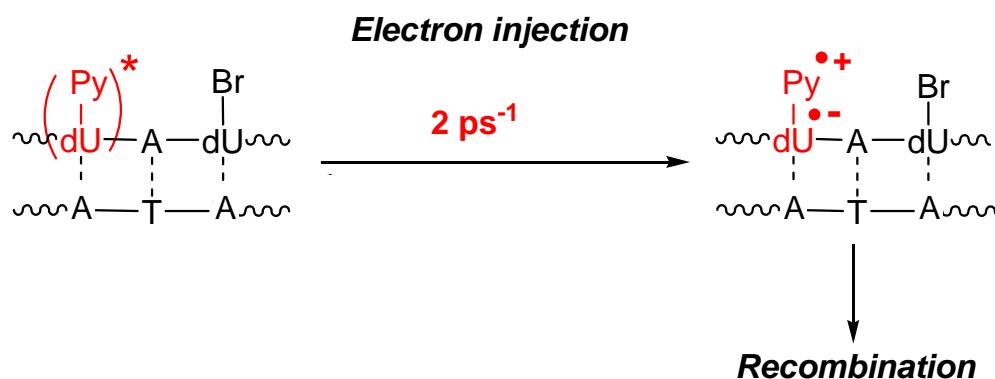
Chem. Eur. J. **2005**, *22*, 1871.

Angew. Chem. Int. Ed. **2005**, *44*, 1636.

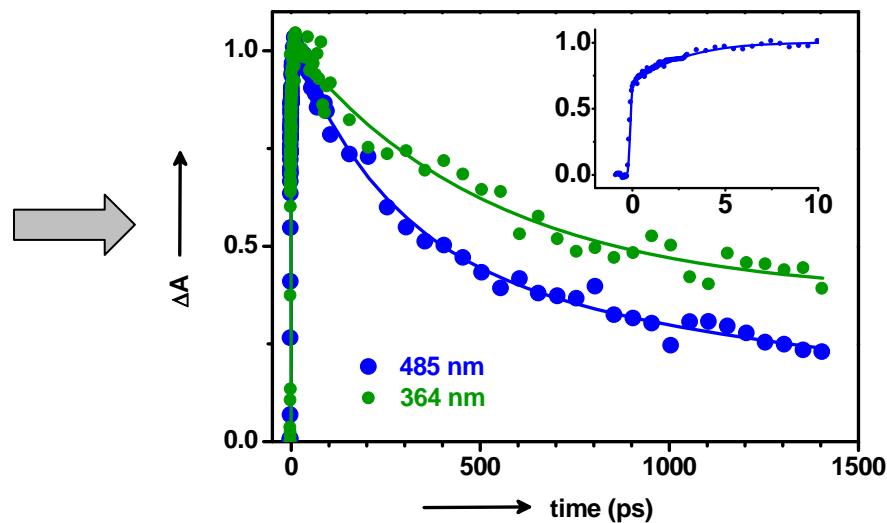
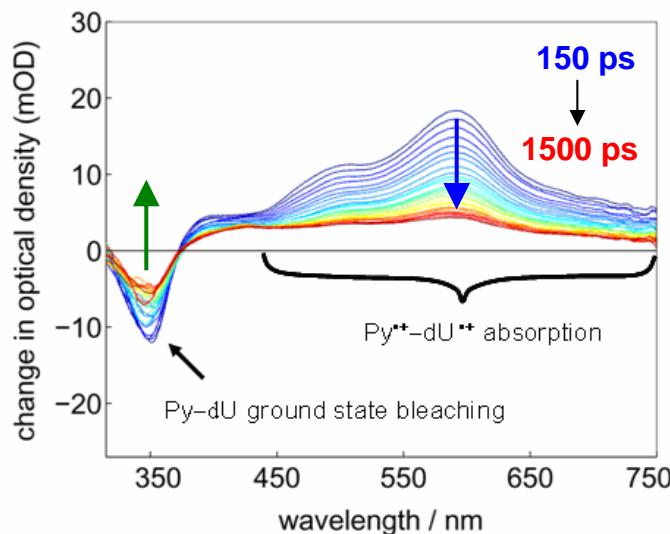
Proc. Natl. Acad. Sci. USA **2006**, *103*, 10192

Part I. Reductive electron transfer in pyrene-modified DNA

Time-resolved spectroscopy

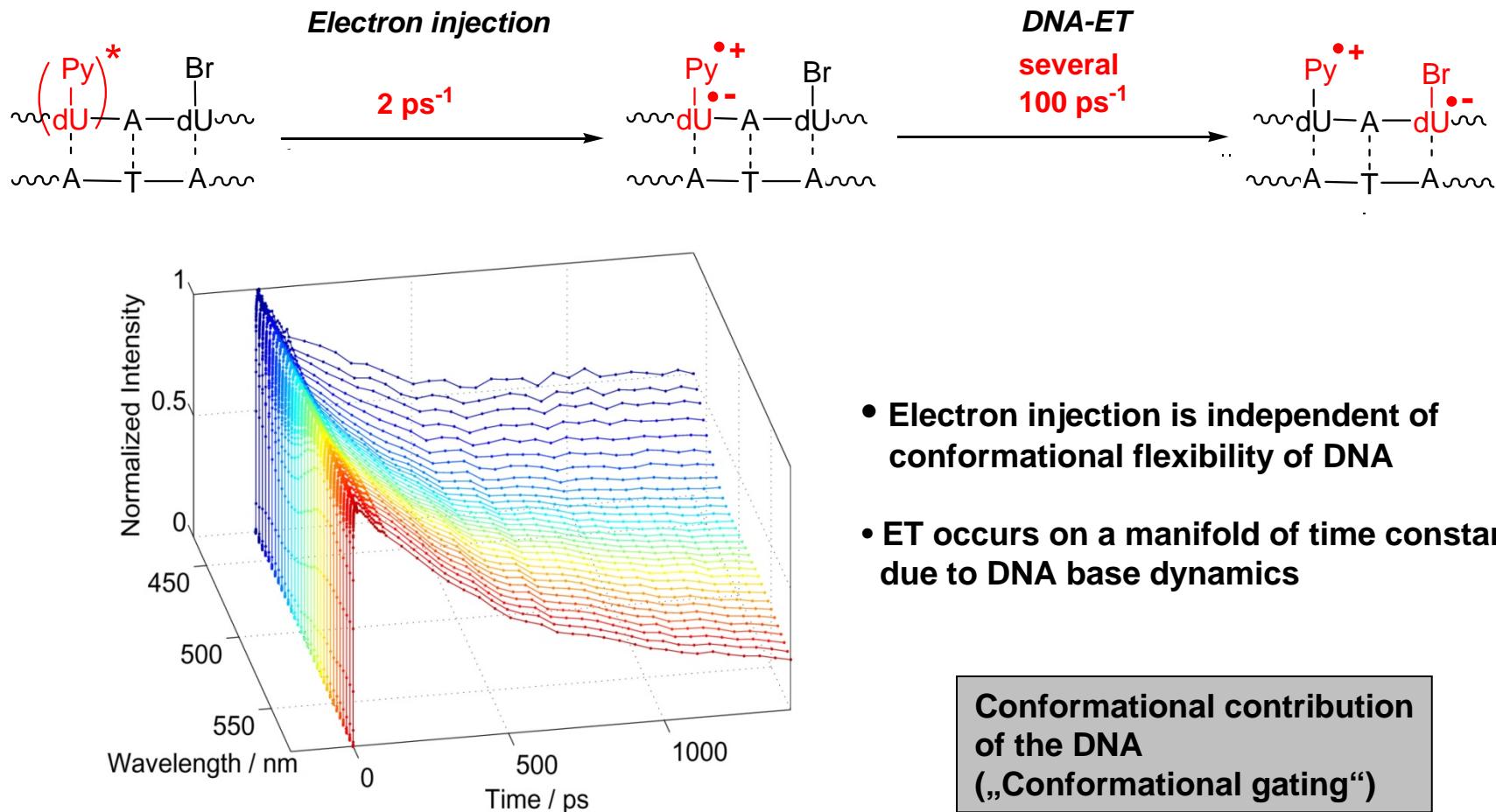


Time-resolved transient absorption spectra



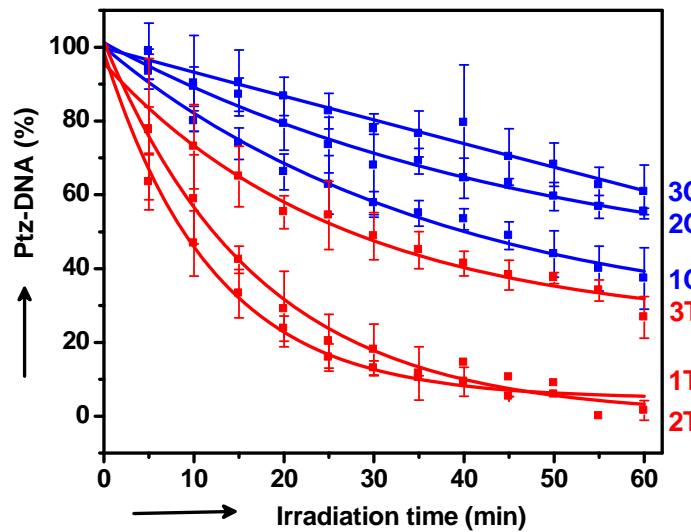
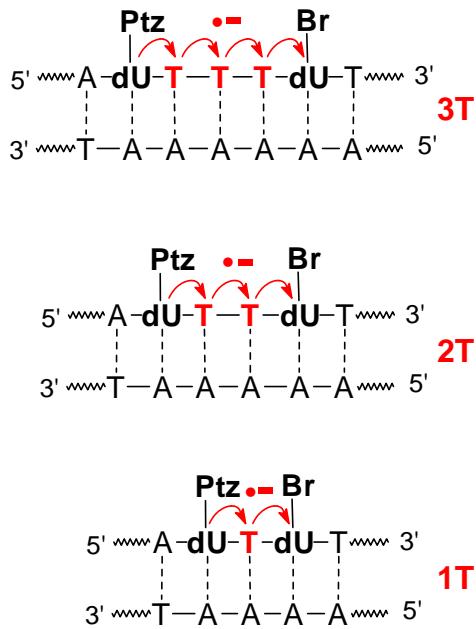
Part I. Reductive electron transfer in pyrene-modified DNA

Influence of DNA dynamics

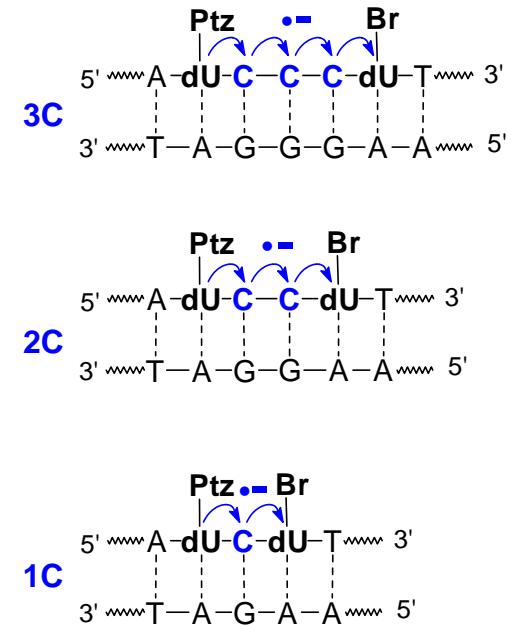


Part I. Reductive electron transfer in phenothiazine-modified DNA

Chemical experiments

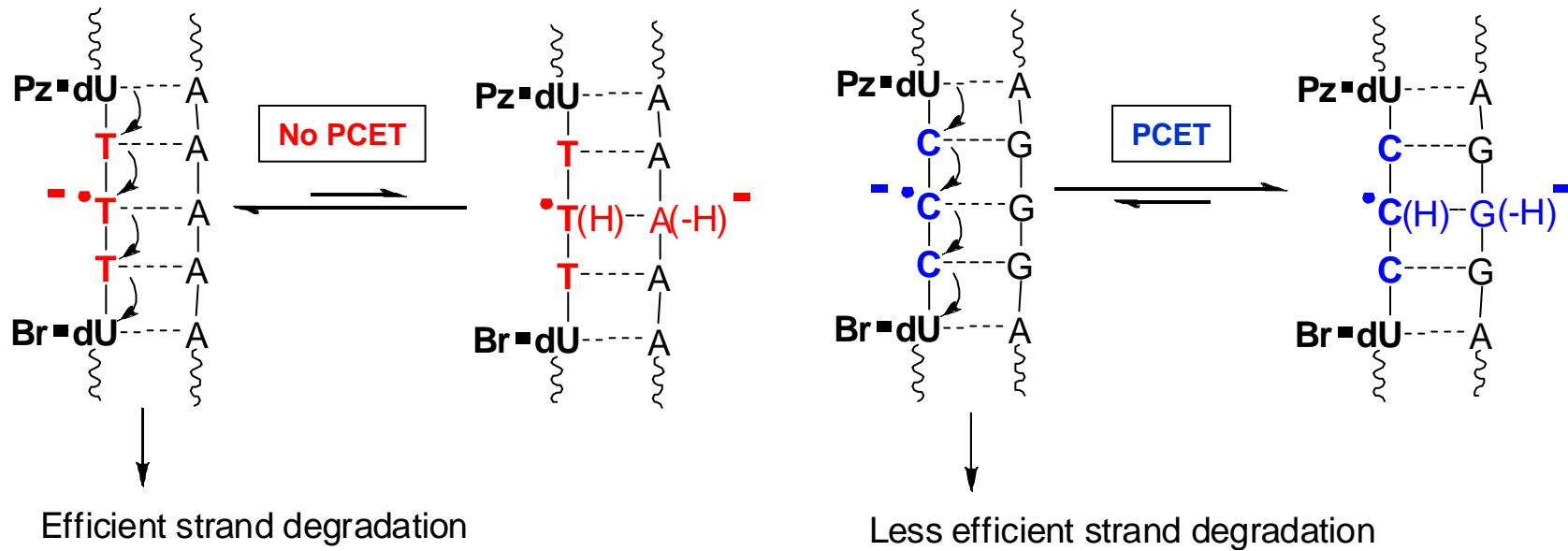


DNA base sequence
and distance dependence



Part I: Reductive electron transfer in DNA

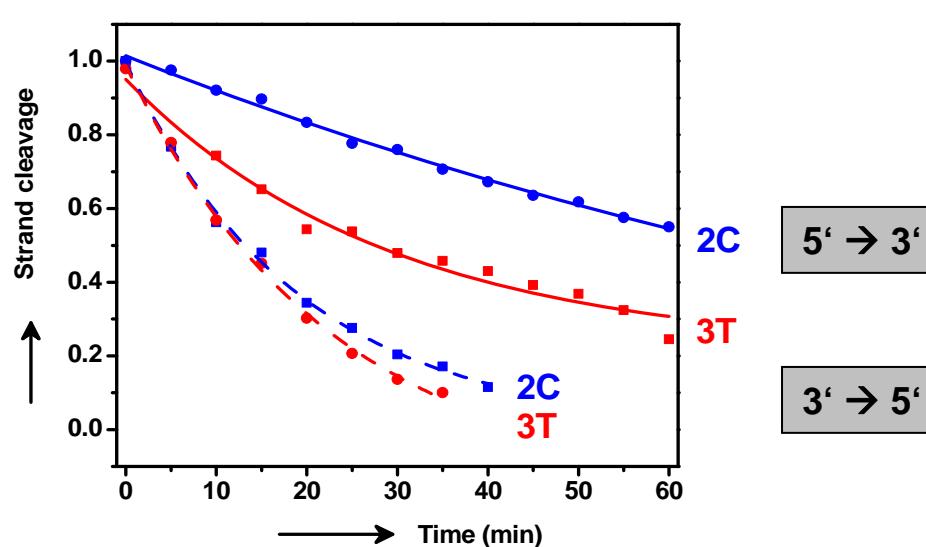
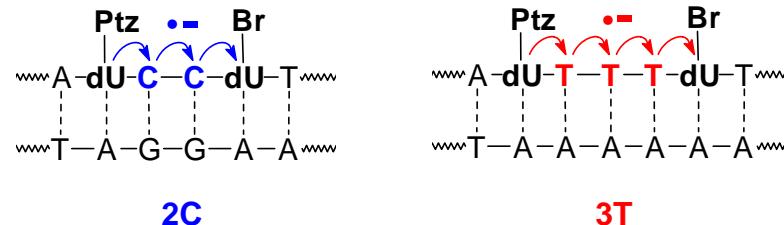
Mechanism of electron hopping



- Each base pair can participate
- Question of long range ET in G-C rich DNA?

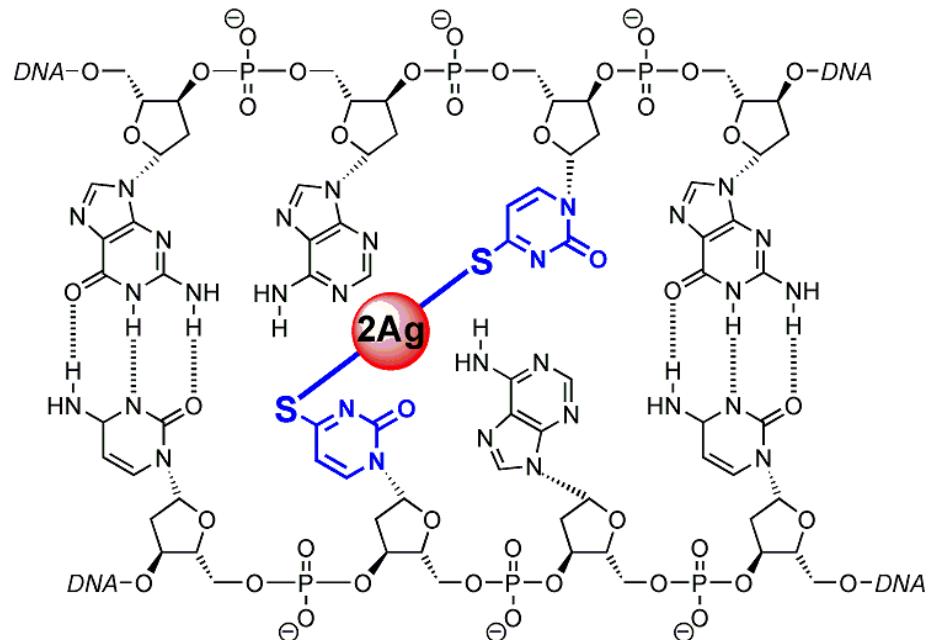
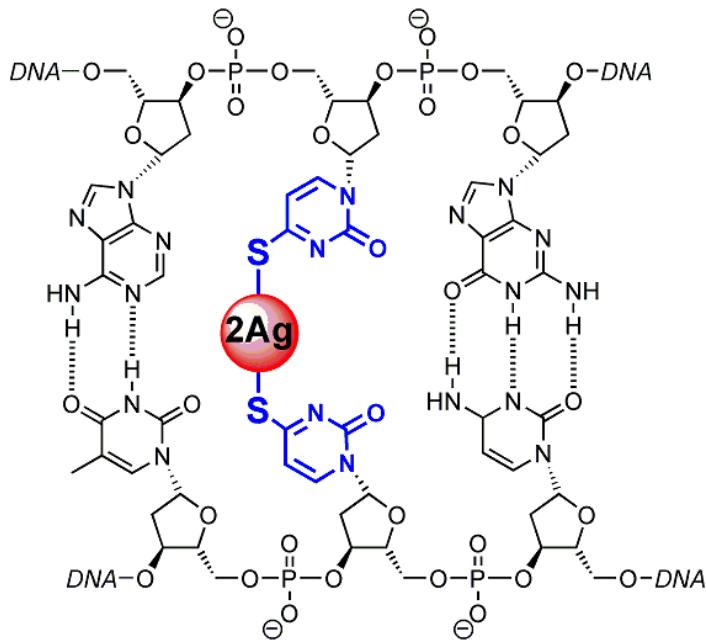
Part I. Reductive electron transfer in phenothiazine-modified DNA

Directionality



Part I. Reductive electron transfer in M-DNA

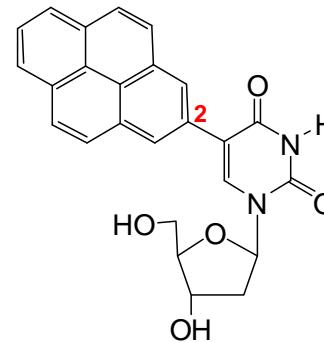
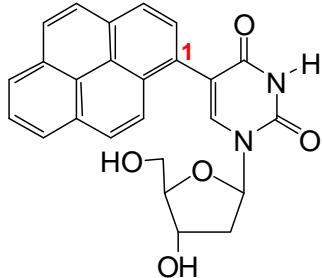
Thio-dU-Ag(I) base pairs



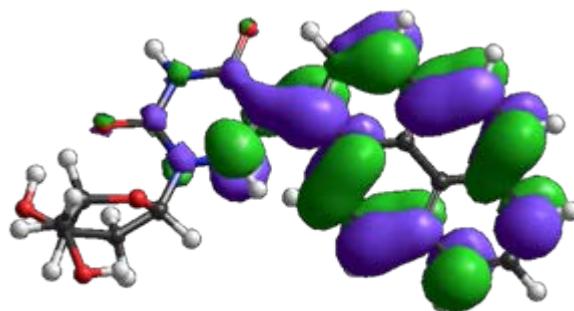
Janez Barbaric

Based on metallated base pair by Simone Peters in the group of Elmar Weinhold, Aachen

Photochemistry of pyrene-modified DNA bases

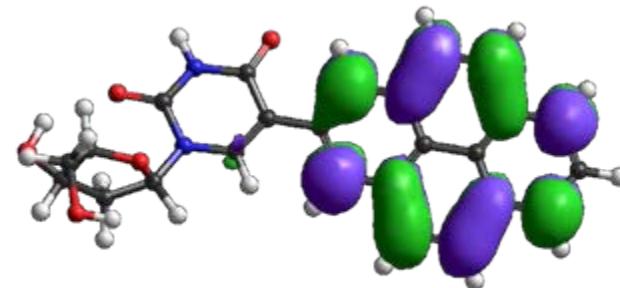


LUMO



Exciplex

$(Py^{\bullet+} dU^{\bullet-})^*$

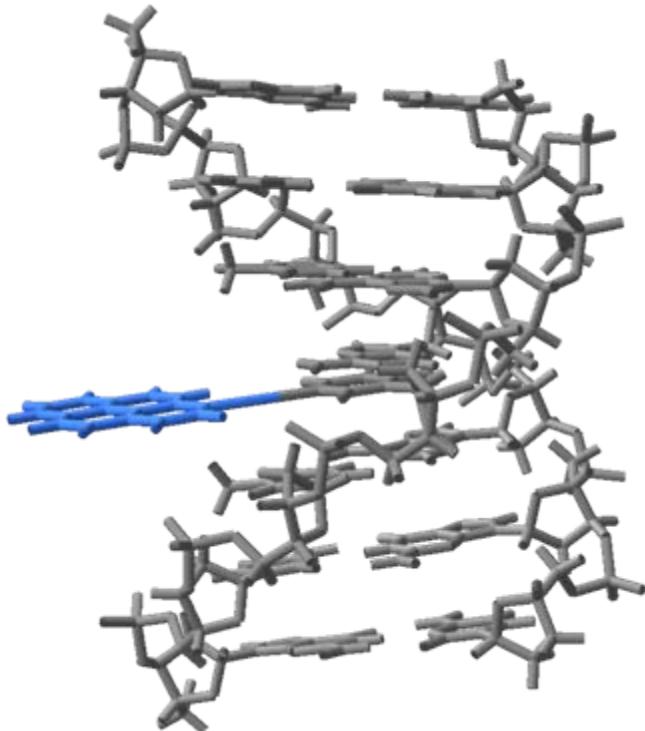


Locally Excited

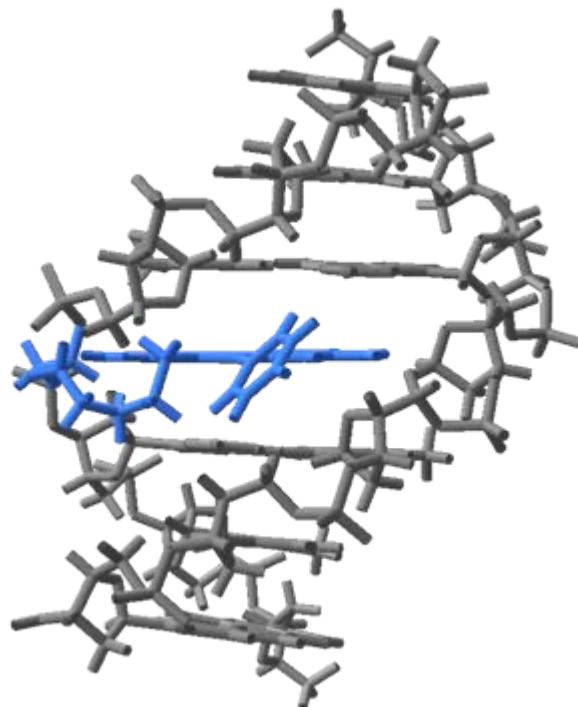
$Py^{\bullet*}-dU$

Part I: Electron donor placement

Functionalization of DNA



DNA base modification



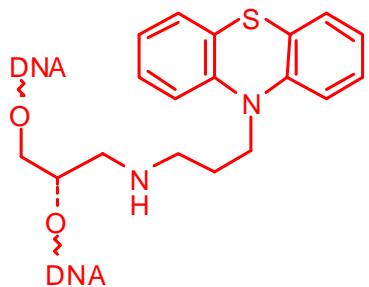
DNA base substitution

Part I. Electron donor placement

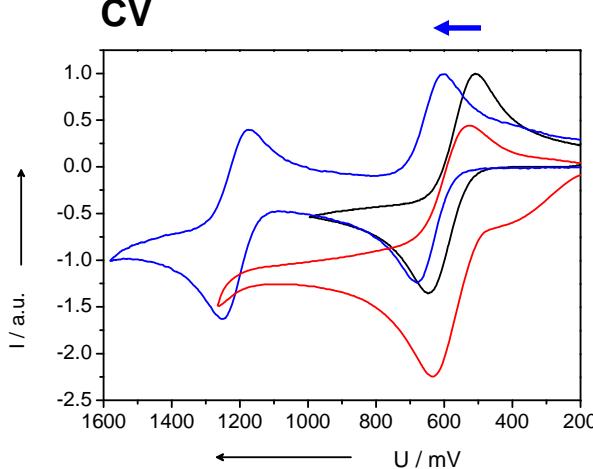


DNA Base substitution vs. base modification: Phenothiazine

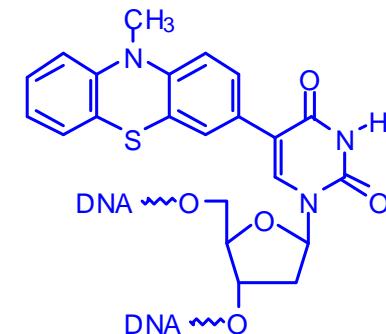
DNA base substitution



CV



DNA base modification

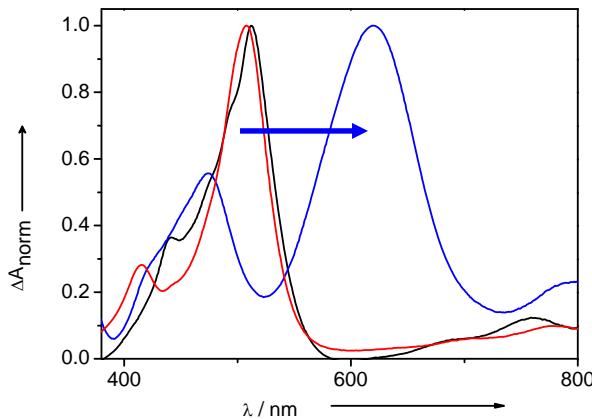


T_m values (17mer DNA)

Counterbase

A	50 °C
C	50 °C
G	50 °C
T	49 °C

Spectroelectrochemistry



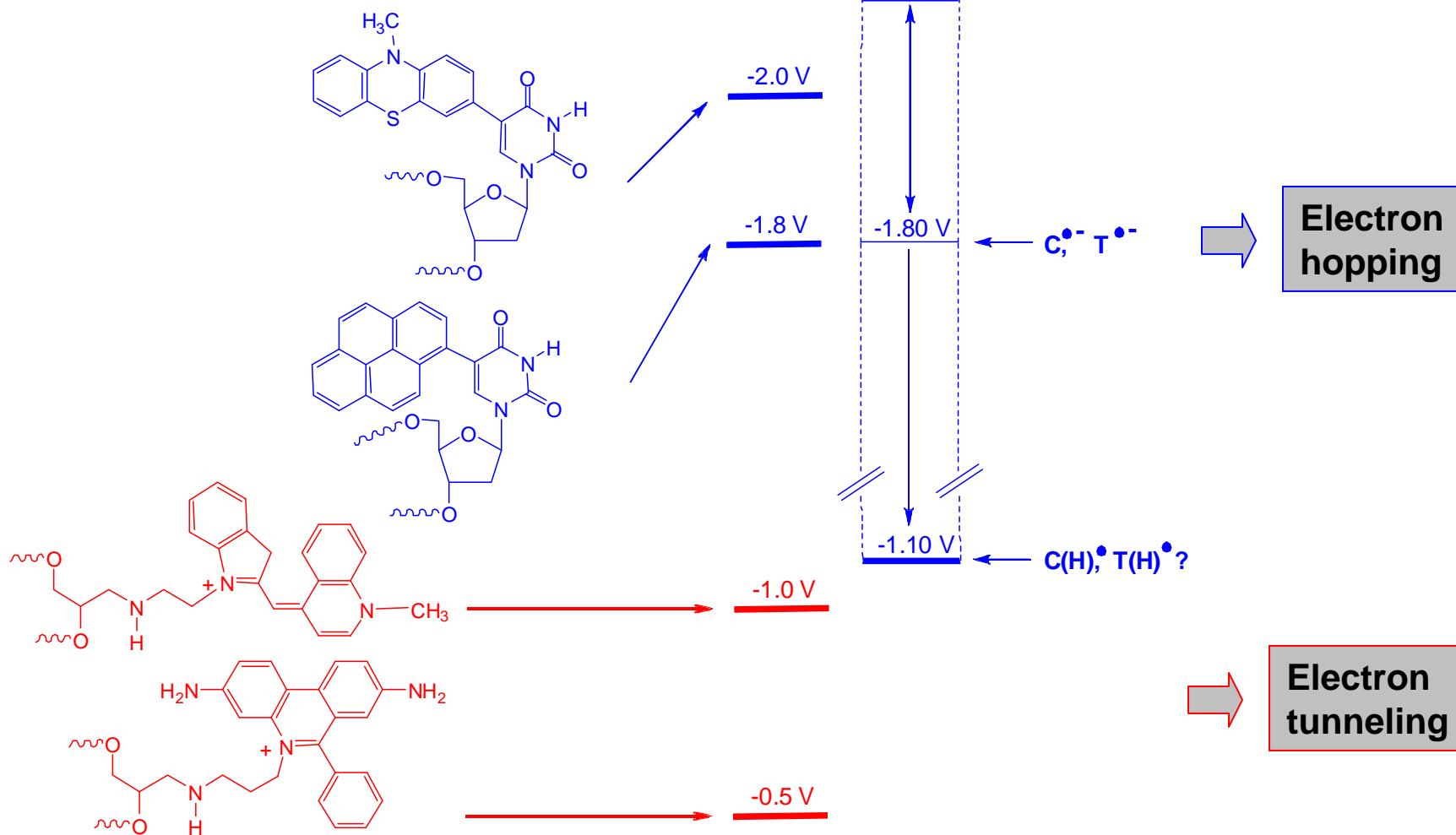
T_m values (17 mer DNA)

Counterbase

A	60 °C
C	56 °C
G	55 °C
T	55 °C

Part II: Photoinduced electron transfer in DNA

Electron donors



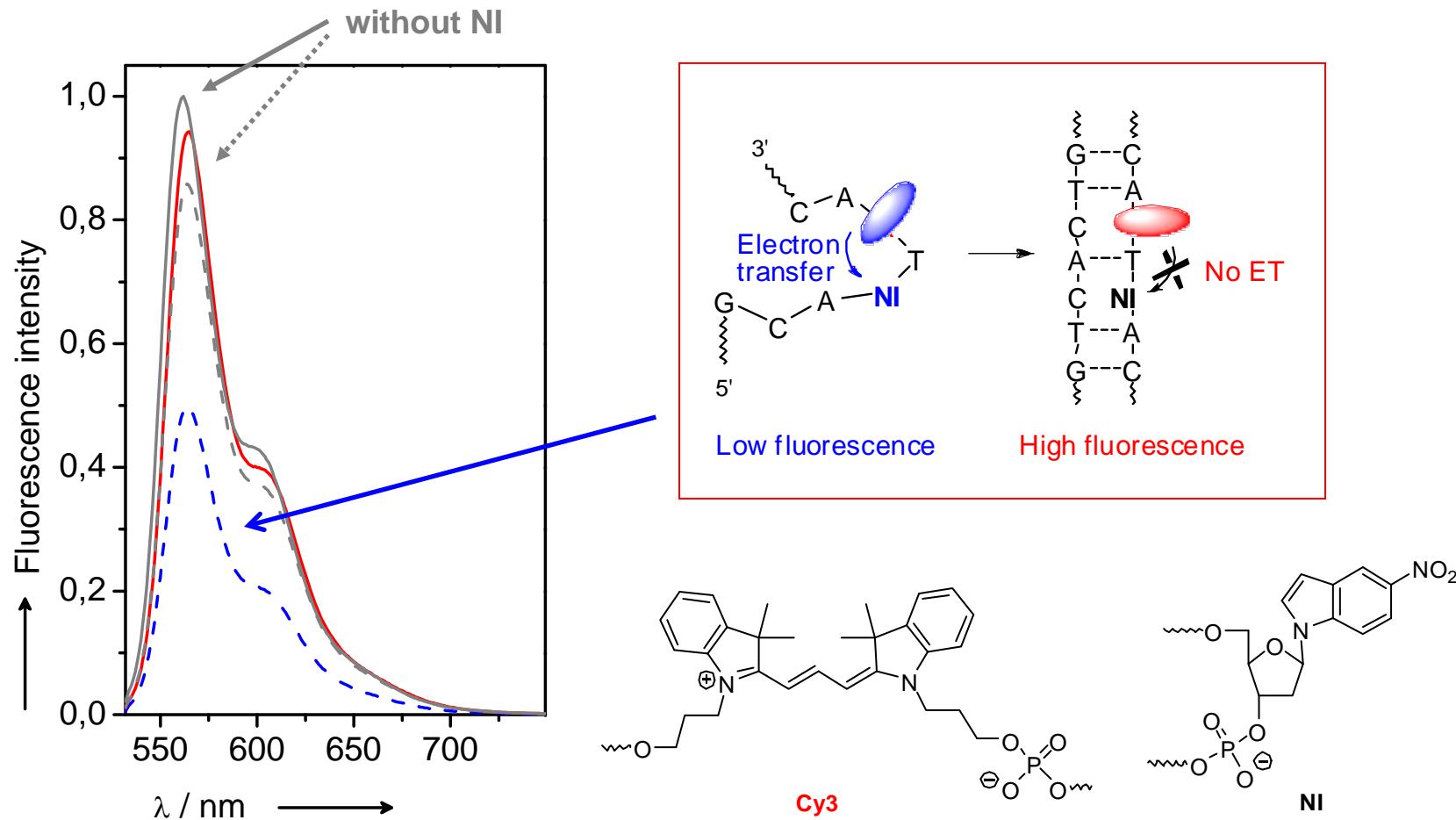
H.-A. Wagenknecht (Ed.), *Charge Transfer in DNA*, Wiley-VCH, 2005, 1.

Angew. Chem. Int. Ed. 2003, 42, 2454; *Curr. Org. Chem.* 2004, 8, 251; *Nat. Prod. Rep.* 2006, 23, 973.

Part II: Charge transfer with cyanines

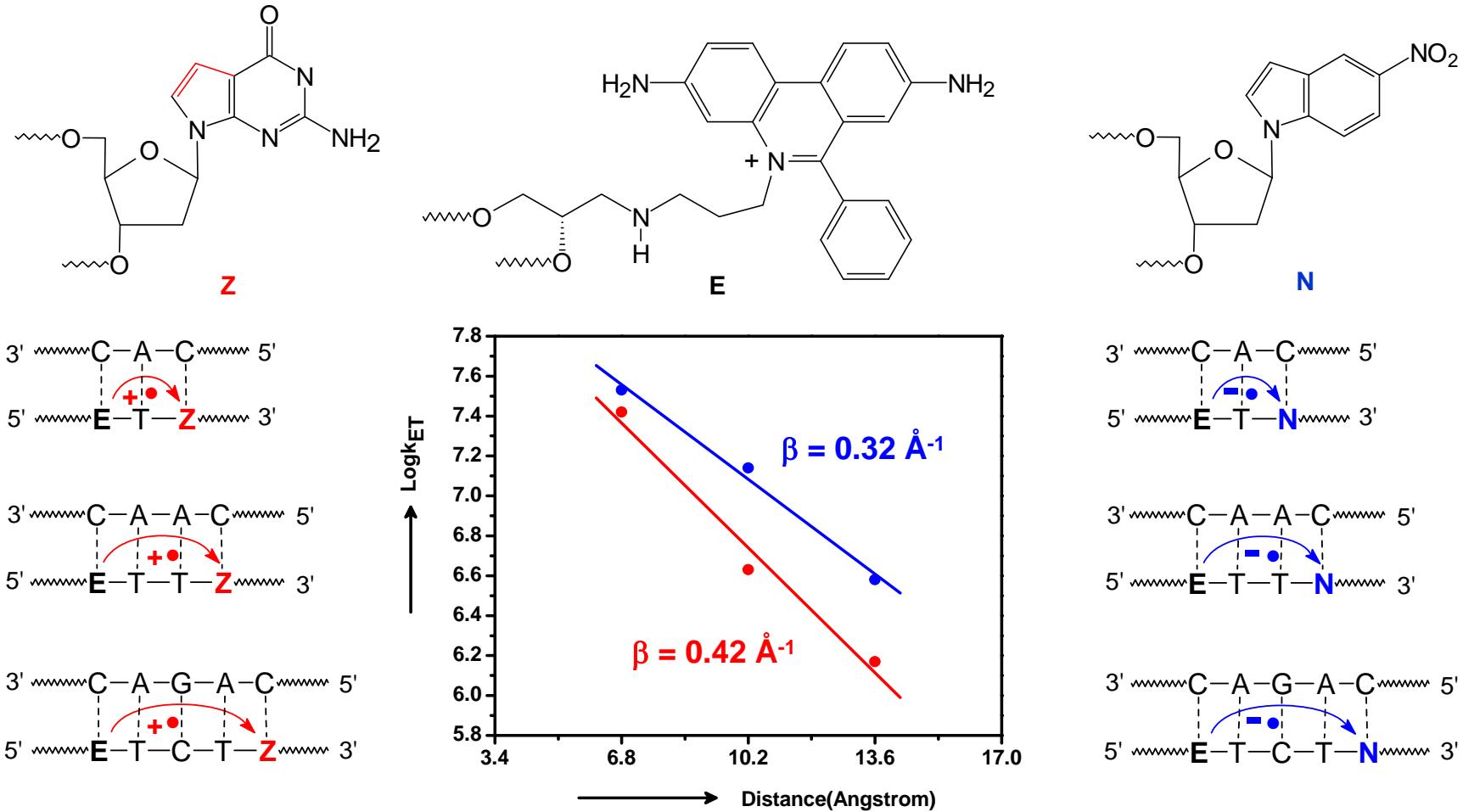


Non-intercalative mode: Cyanine dyes as the charge donor



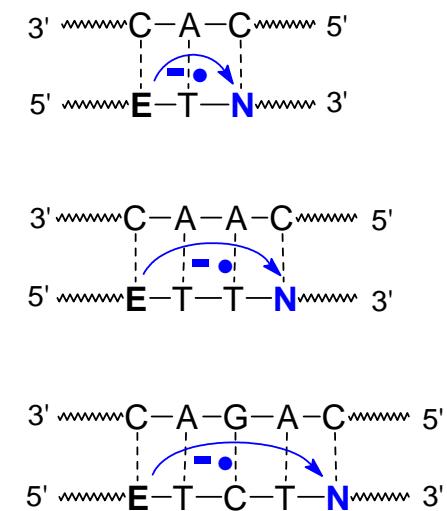
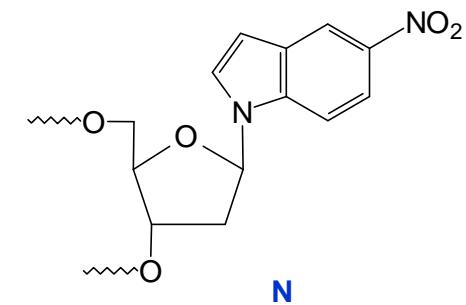
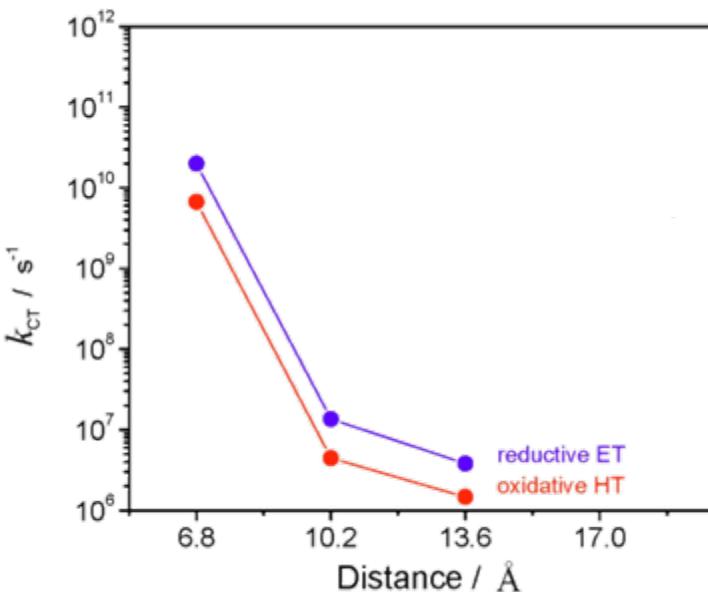
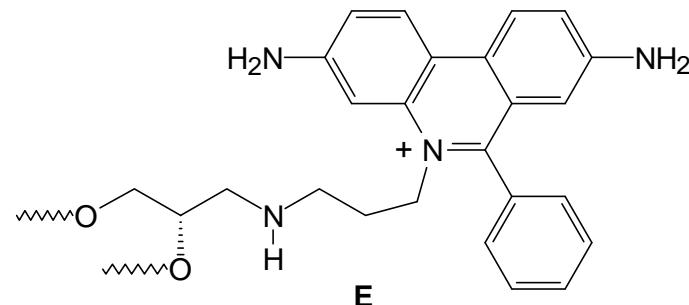
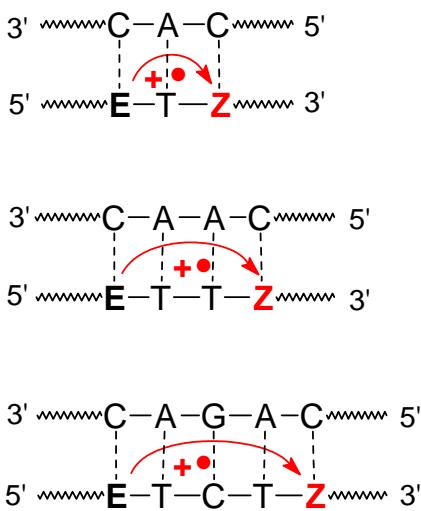
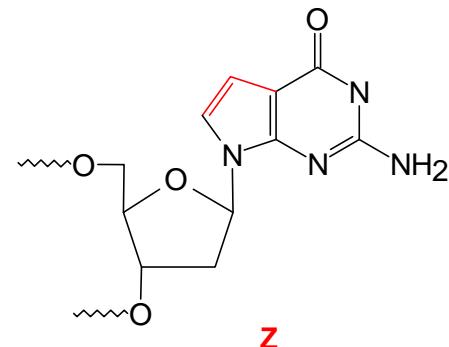
Part II: Charge transfer in ethidium-modified DNA

Hole vs. electron transfer with ethidium



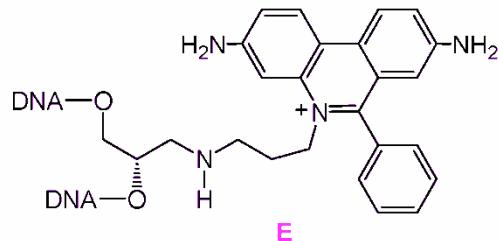
Part II: Charge transfer in ethidium-modified DNA

Intercalative mode: Ethidium as a charge donor

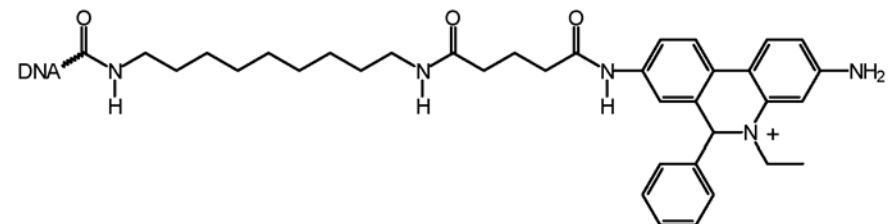


Part II: Charge transfer in ethidium-modified DNA

Conformational gating

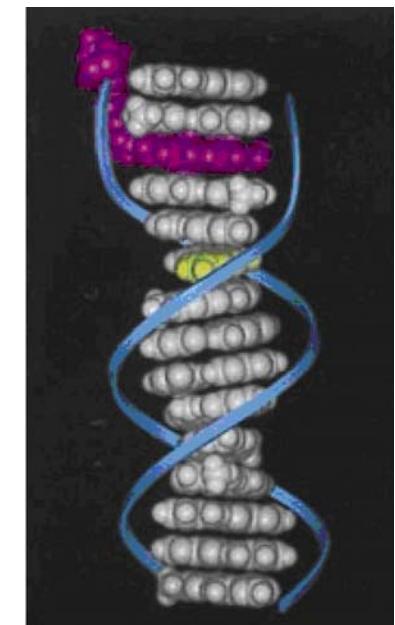
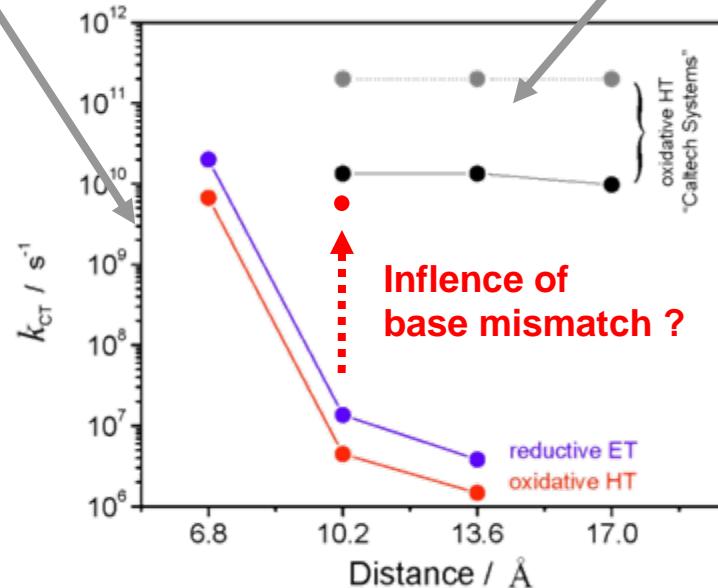
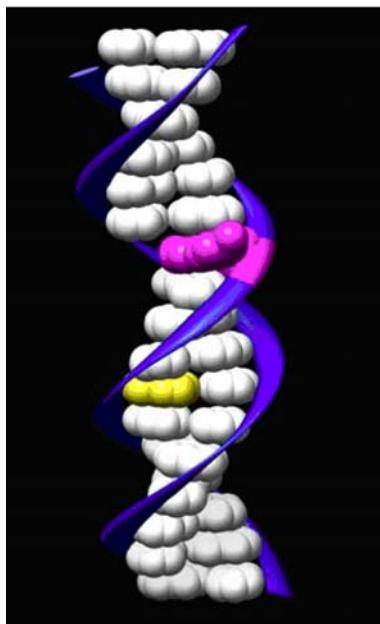


Ethidium base pair surrogate



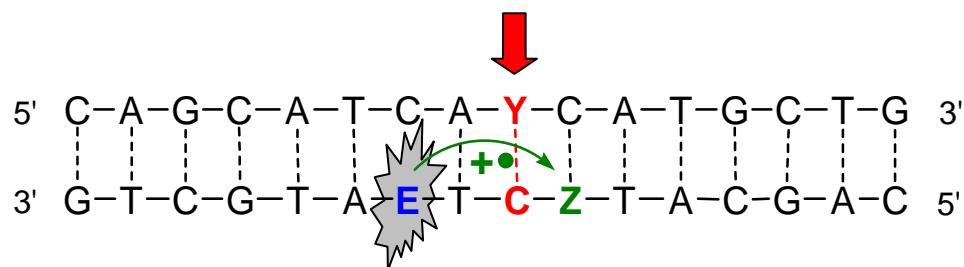
rigid vs. flexible

Ethidium linker in the "Caltech Systems"

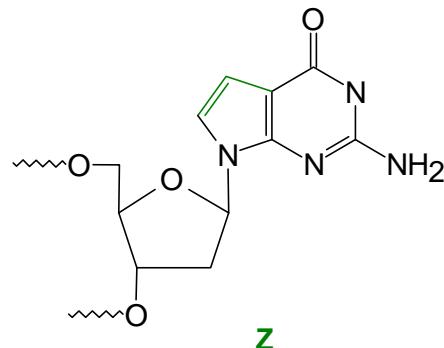
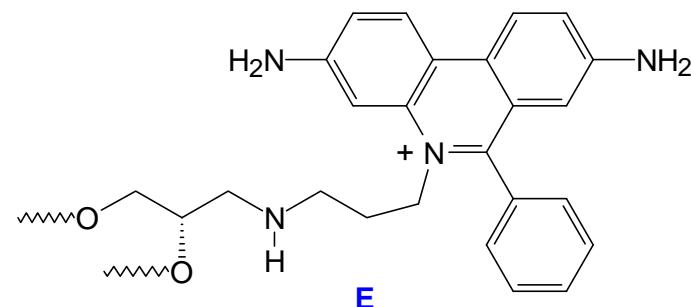
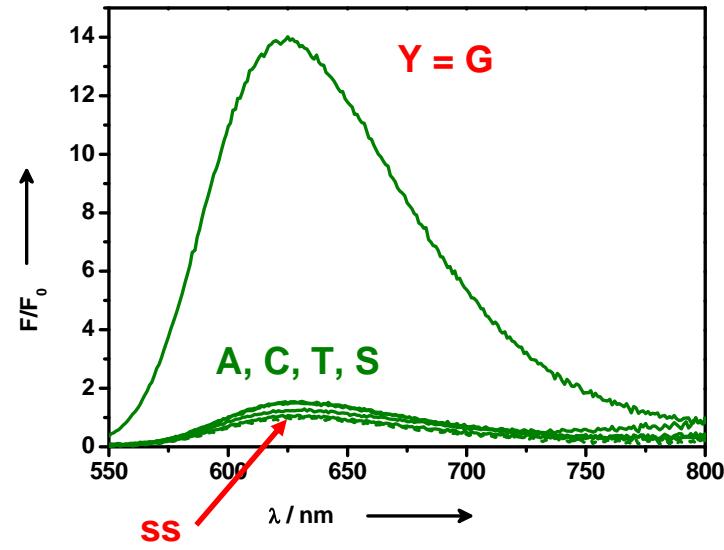


Part II. Charge transfer in ethidium-modified DNA

Single base mismatch detection



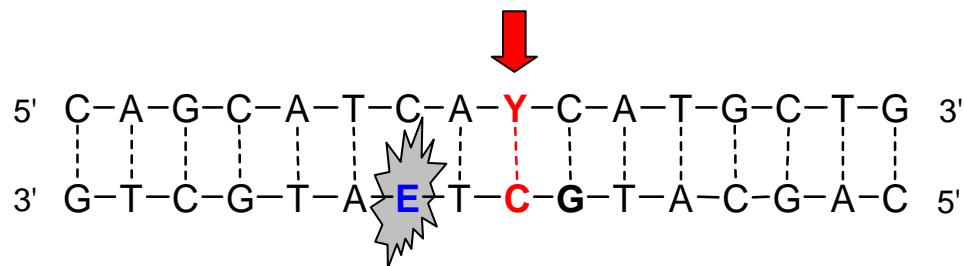
With charge transfer



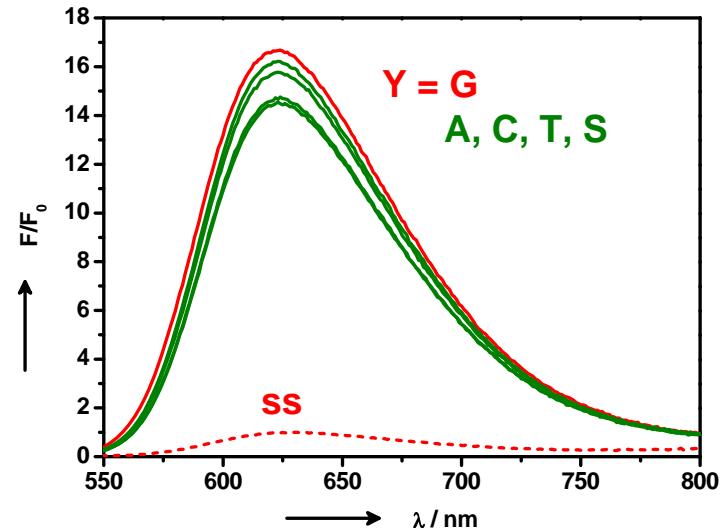
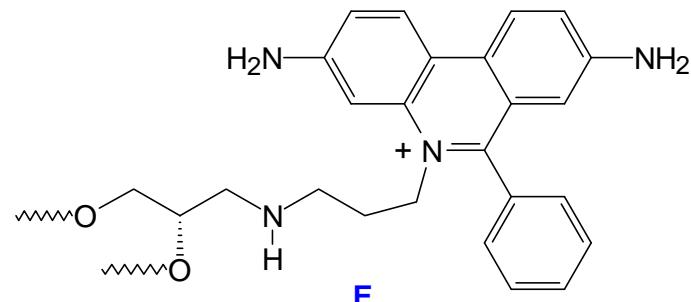
Strong
match/mismatch
discrimination

Part II. Charge transfer in ethidium-modified DNA

Single base mismatch detection

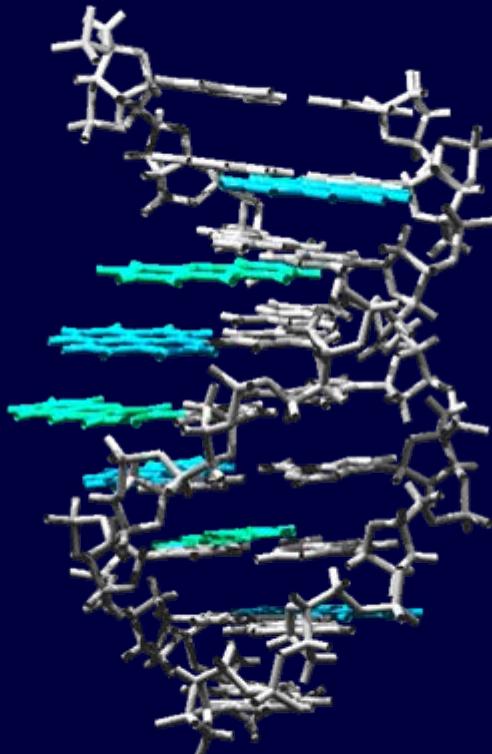
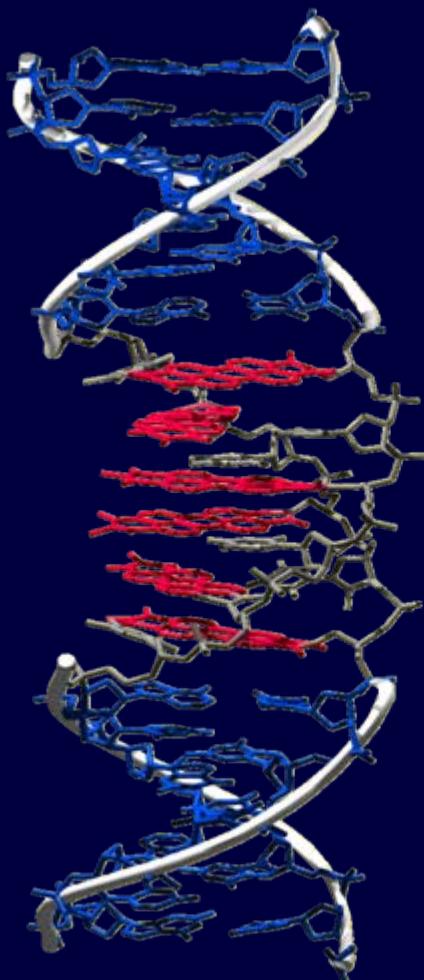


Control: without charge transfer



No significant
match/mismatch
discrimination

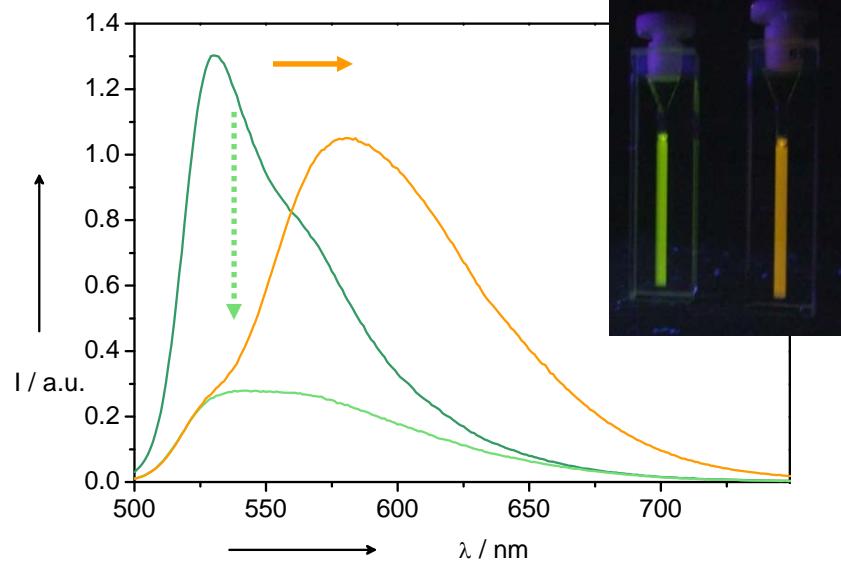
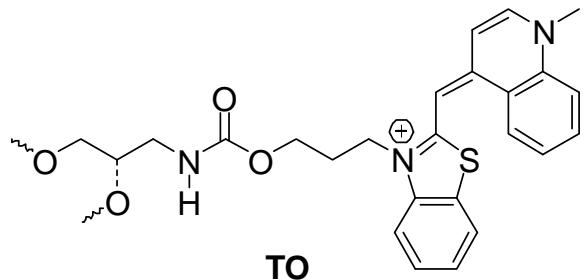
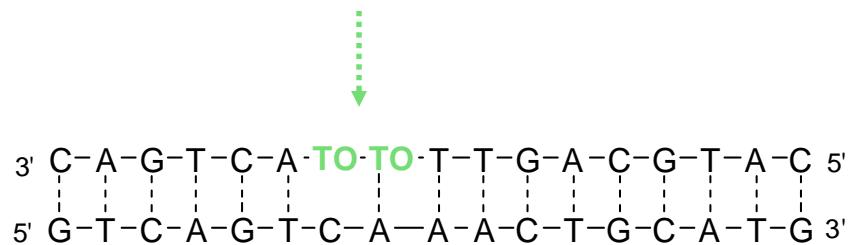
Part III: DNA architectures for switchable optical properties



Part III: DNA base substitutions



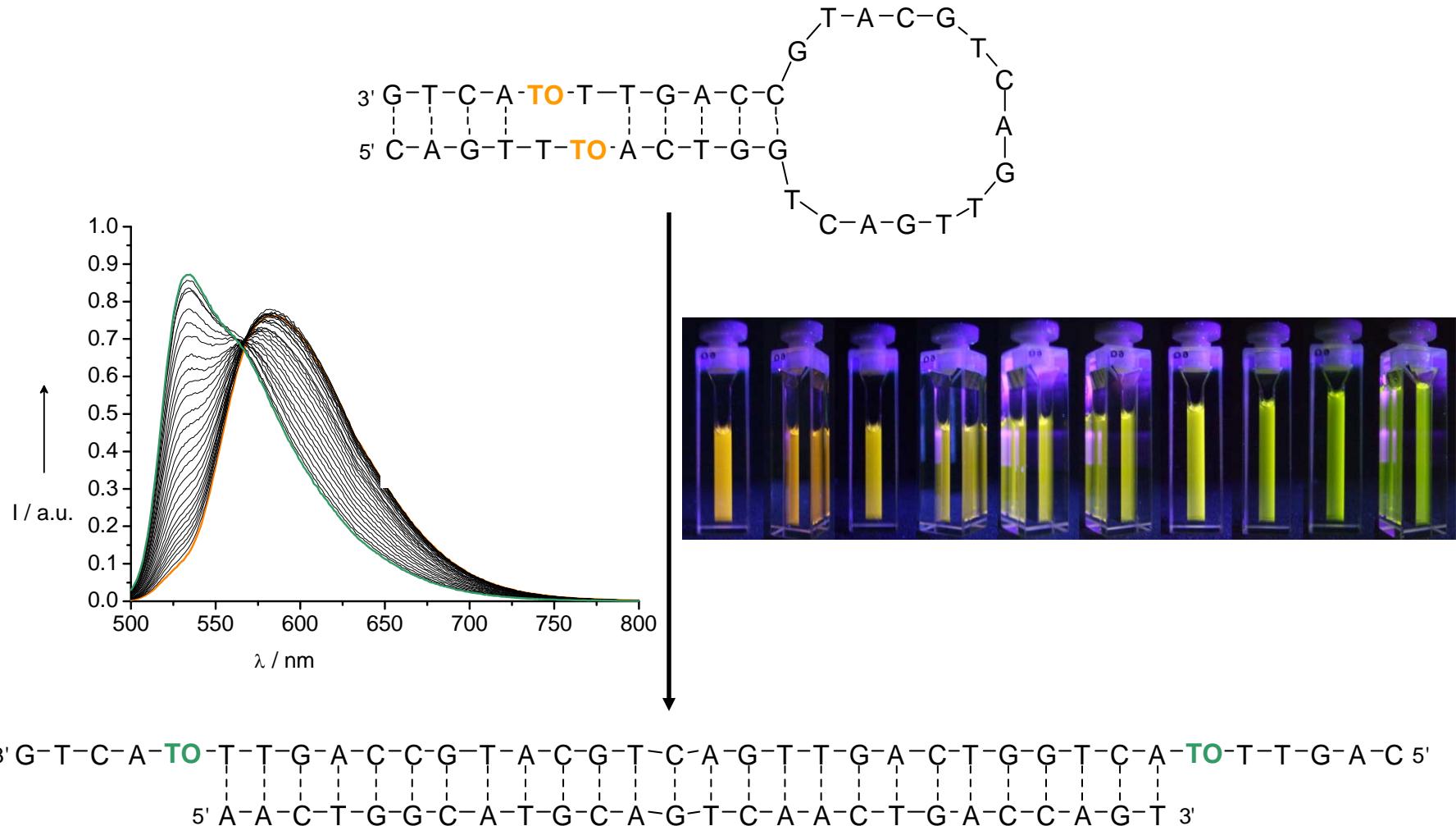
Interstrand thiazole orange dimers



Part III: DNA base substitutions



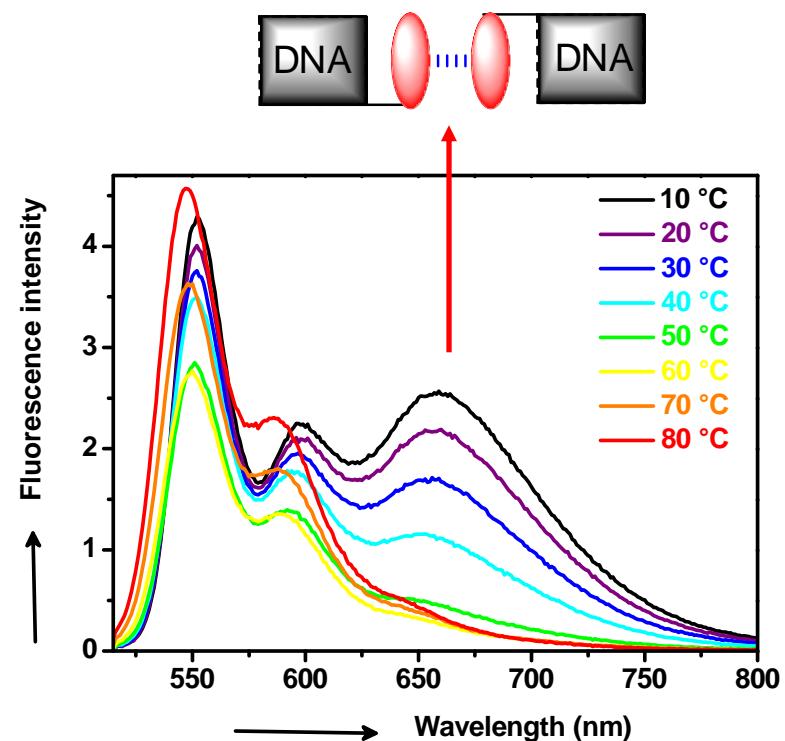
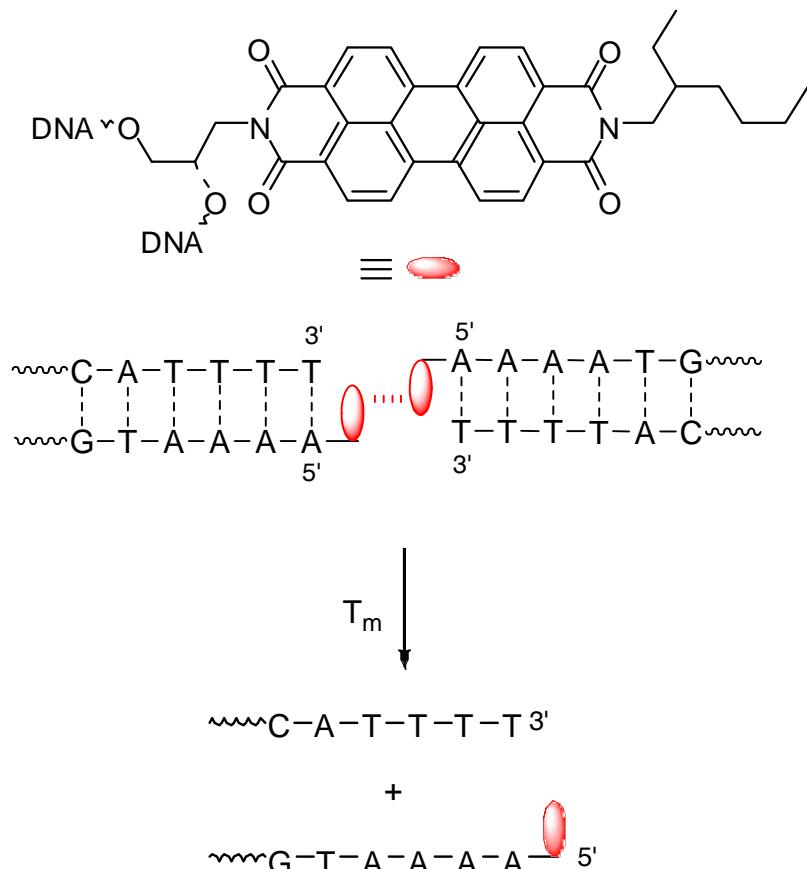
Interstrand thiazole orange excimers



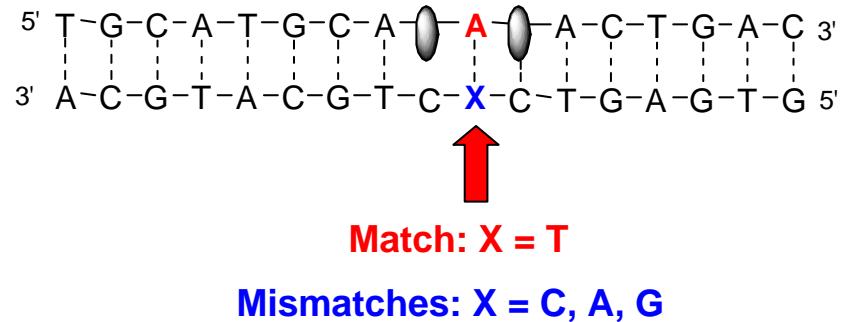
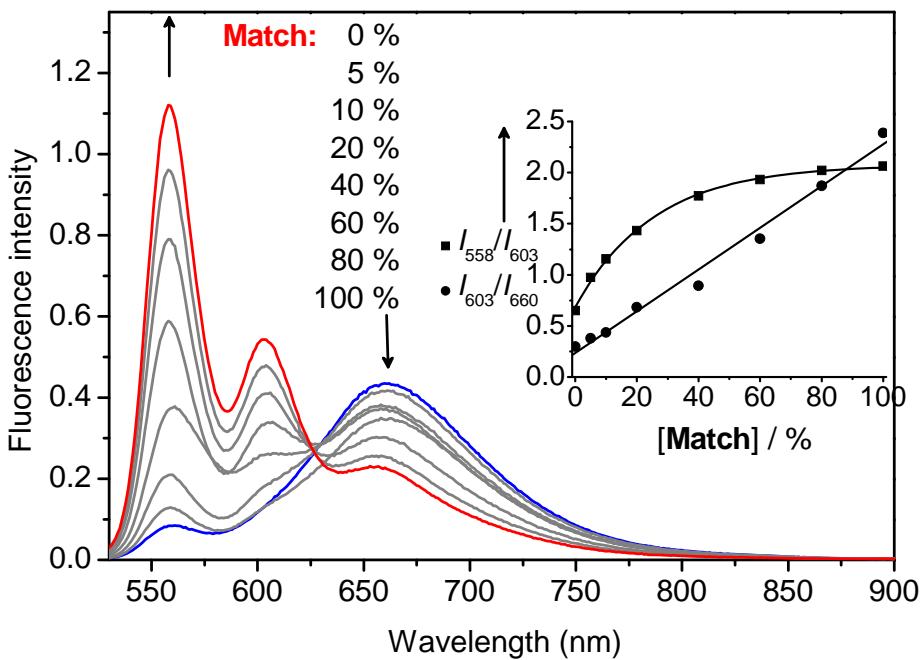
Part III. DNA base substitutions



Aggregation of perylenebisimide-capped DNA



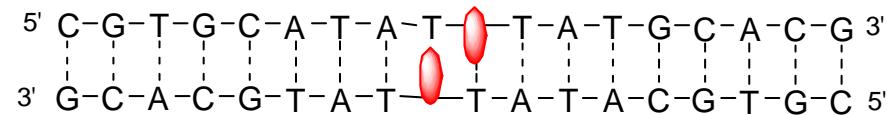
Interstrand perylenebisimide dimers



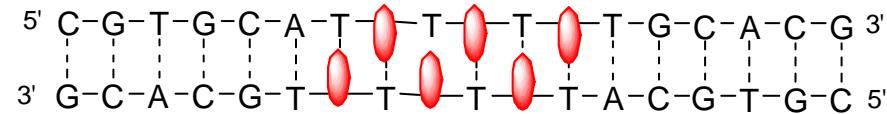
Part III: DNA base substitutions



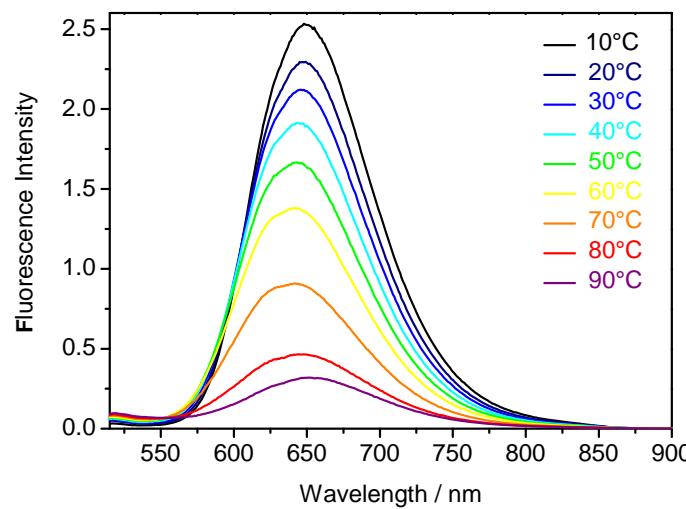
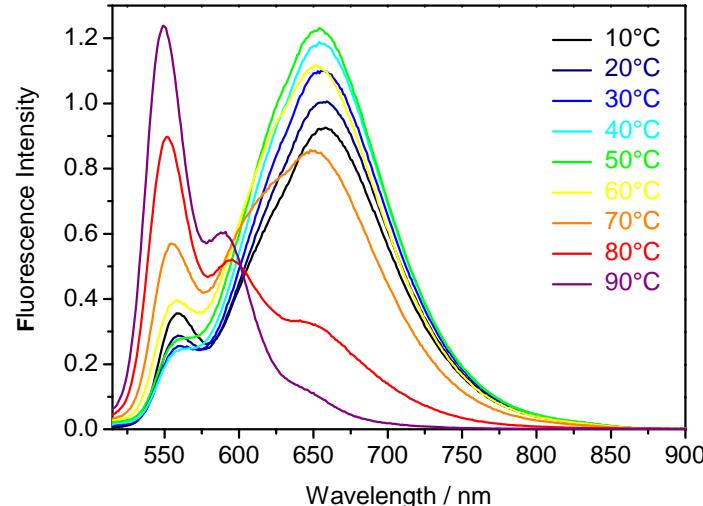
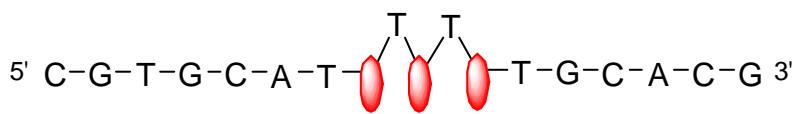
Interstrand perylenebisimide-zippers



ΔT



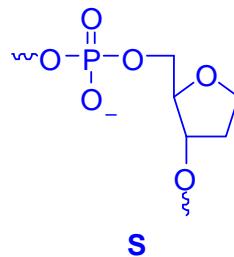
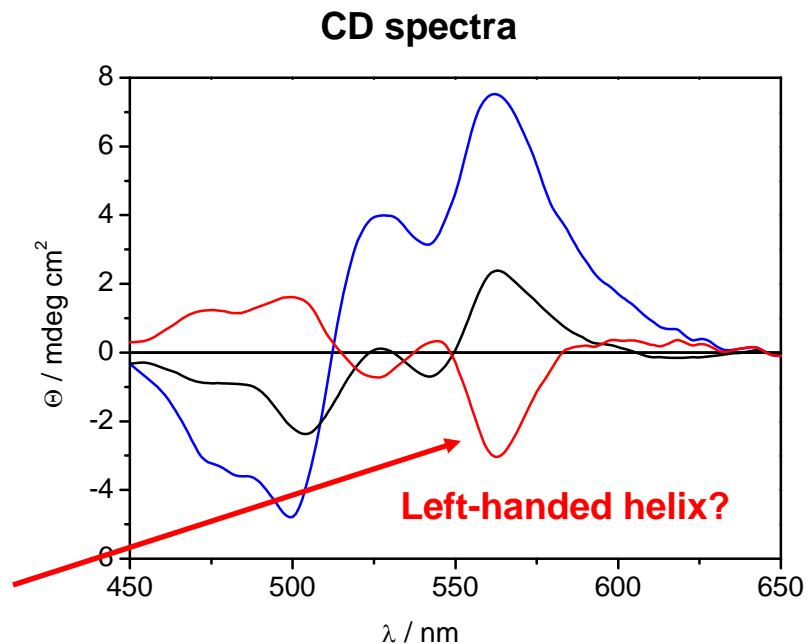
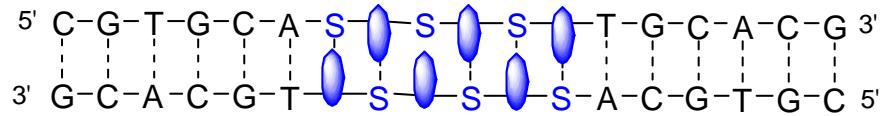
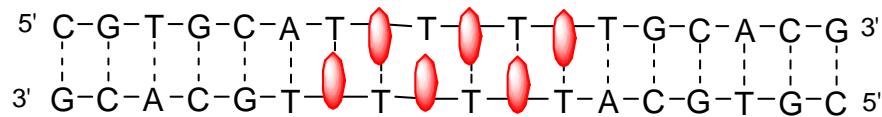
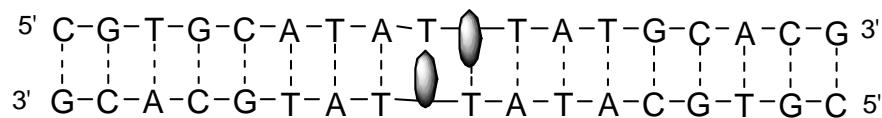
ΔT



Part III: DNA base substitutions



Interstrand perylenebisimide-zippers

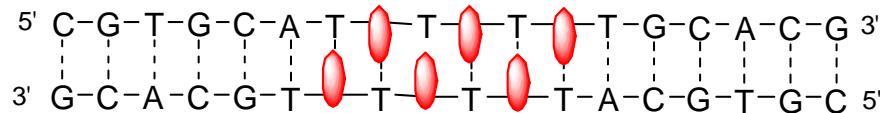
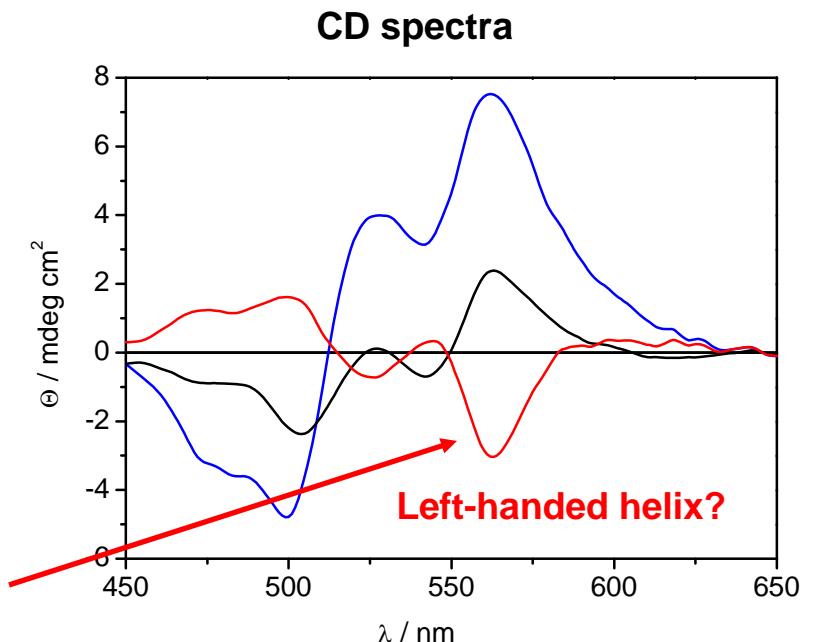
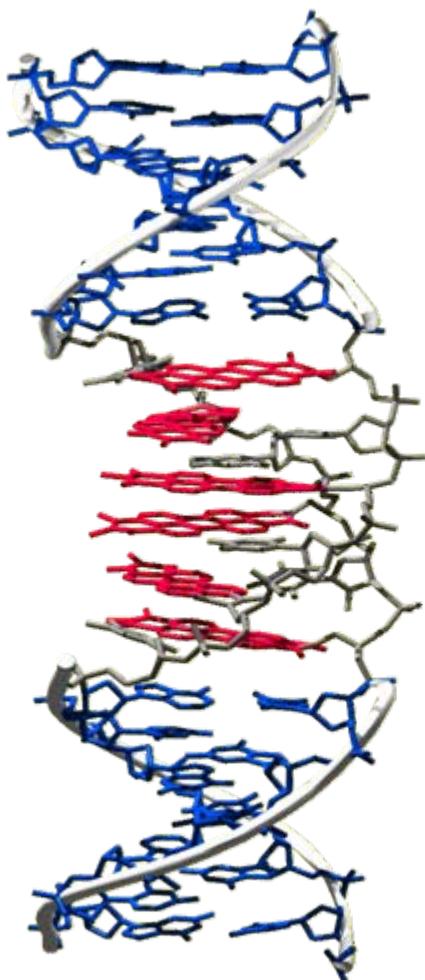


Part III: DNA base substitutions



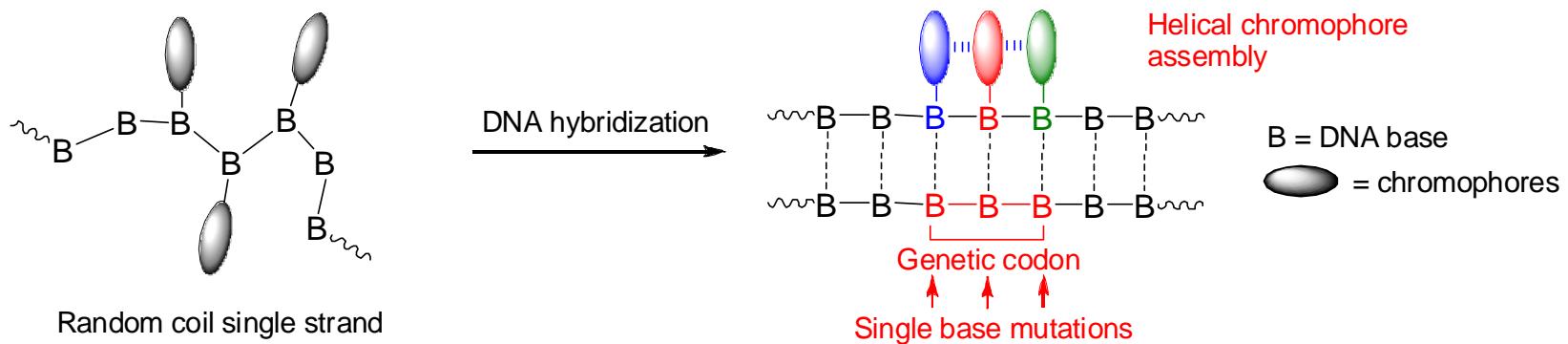
Interstrand perylenebisimide-zippers

35-45 ° left
85-95 °C right
35-45 ° left
85-95 °C right
35-45 ° left



$$\Delta\epsilon = f(\sin(2\lambda))$$

Multifluorophores based on DNA base modifications

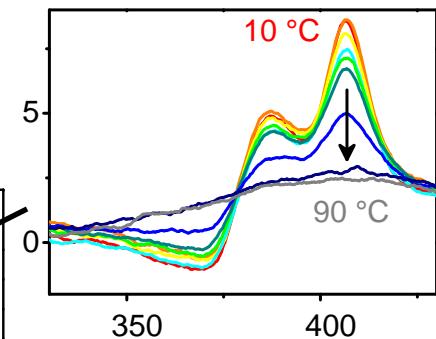
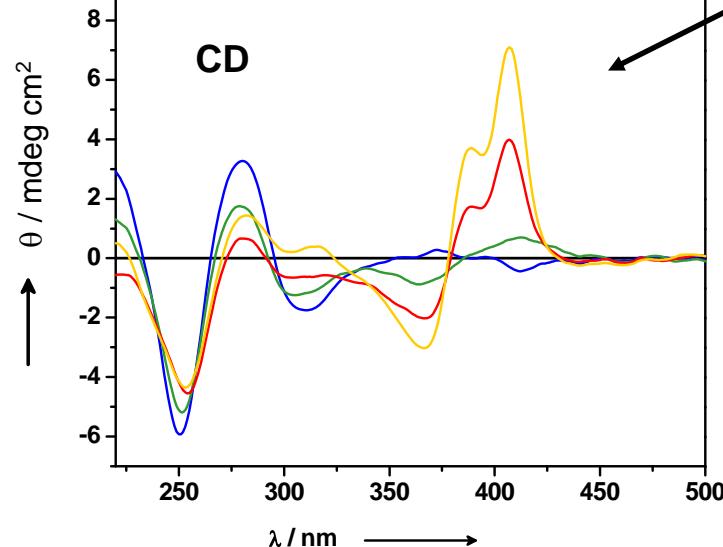
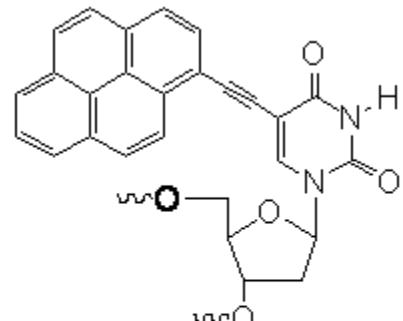
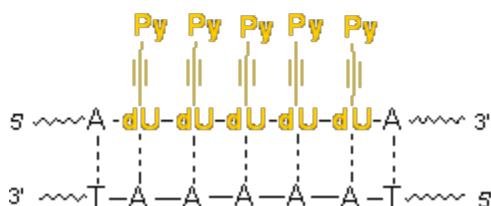
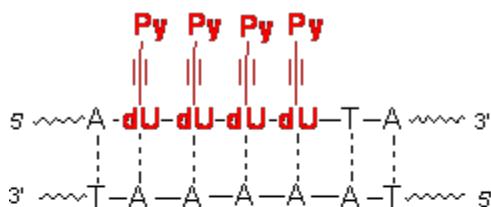
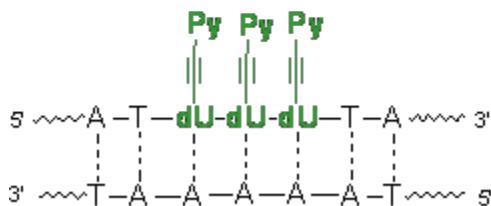
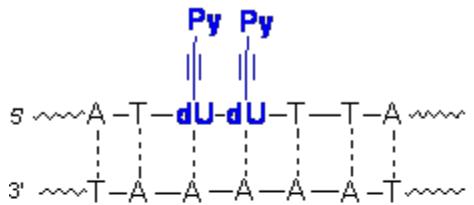


together with fs-resolved microarray readout

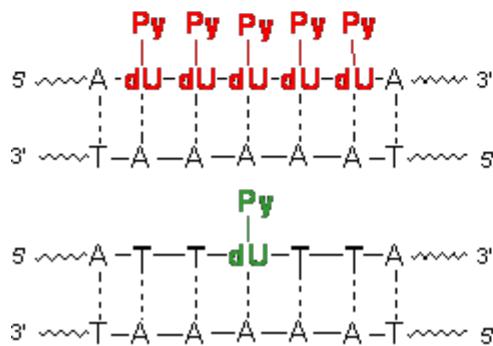
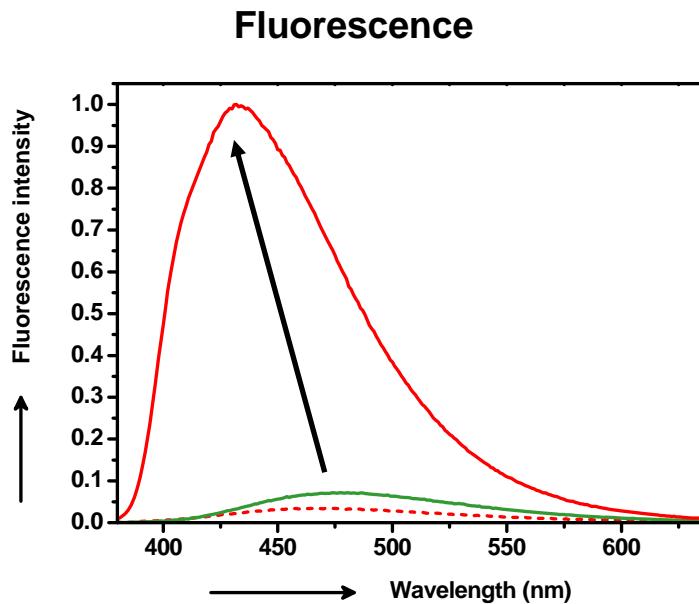
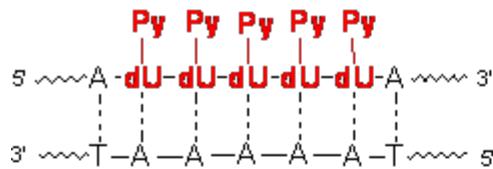
Part III: DNA base modifications



Multiple Py-≡-dU-labels

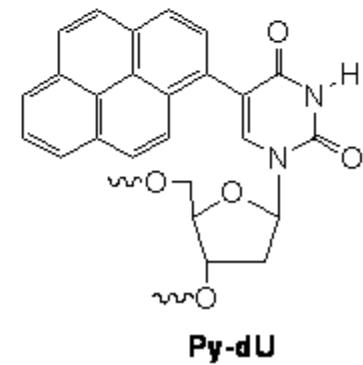


Multiple Py-dU-labels



ds/ss = 22

PydU₅/PydU₁ = 11



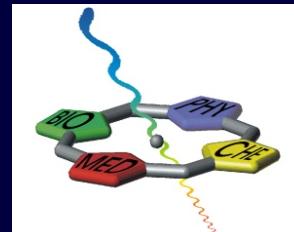


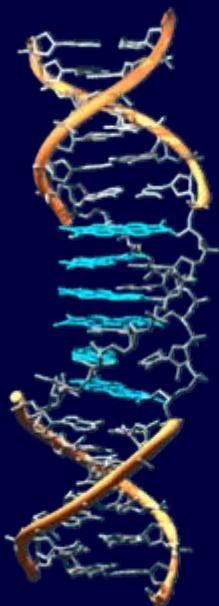
Vielberth-Symposium on Functional Nucleic Acids

(IV. Nucleinsäurechemie-Treffen)

10. – 11. September 2009

University of Regensburg (Germany)





Lectures:

Hiroyuki Asanuma (Nagoya, JP)

Shankar Balasubramanian (Cambridge, UK)

Tom Brown (Southampton, UK)

Alexander Deiters (Raleigh, North Carolina, USA)

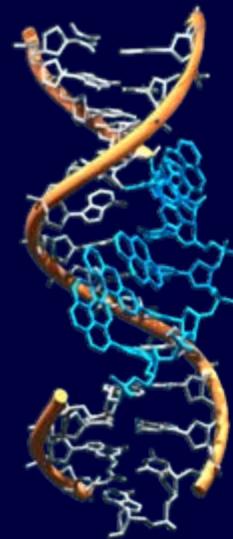
Alexander Heckel (Frankfurt, D)

Christian Leumann (Bern, CH)

Jens Müller (Münster, D)

Floyd Romesberg (Scripps La Jolla, USA)

Poster session including short poster talks



Acknowledgements

